

Poster 32

Optimization of a protocol for isolation of immune cells from European seabass (Dicentrarchus labrax)

D. Salazar-Gutierrez ^{1,2}, M. Ferreira ^{1,2}, V. Sousa ^{1,2}, L. M. P. Valente ^{1,2}, A. Correia ^{2,3,4} and <u>S. Gomes ^{1,2,*}</u>

¹ CIIMAR/CIMAR-LA, Centro Interdisciplinar de Investigação Marinha e Ambiental, 4450-208 Matosinhos, Portugal

² ICBAS, Instituto de Ciências Biomédicas de Abel Salazar, Universidade do Porto, 4050-313 Porto, Portugal

³ i3S, Instituto de Investigação e Inovação em Saúde, Universidade do Porto, 4200-135 Porto, Portugal

⁴ IBMC, Instituto de Biologia Molecular e Celular, Universidade do Porto, 4150-180 Porto, Portugal

* Correspondence: sagomes@icbas.up.pt

Abstract

Background: As one of the most important food production sectors worldwide, aquaculture is now invested in developing tools to study fish immunity. Fish health relies heavily on hematopoietic organs (head kidney), but also on intestine. The wide knowledge-gap of the fish immune system coupled with the lack of cell lines to perform *in vitro* studies poses a barrier to the development of efficient tools to enhance fish immunity. Objective: We optimized the protocol for isolation of immune cells from the head kidney (HK) and posterior intestine (PI) of European seabass, to be used as an in vitro tool to test the impact of environmental contaminants on the fish immune system. Methods: HK and PI of European seabass were collected following the methodology of Park et al. [1]. Tissues were pushed through cell strainers. Cell suspensions were layered on a Percoll gradient of 34%/51% (HK) and 25%/75% (PI) and the intermediate band was thereafter collected. Isolated cells were resuspended in L-15⁺ media with 10% FBS and allowed to adhere to cell culture dishes for 24-h at 23°C. Adherent cells were detached with PBS 7 mM EDTA on ice. Isolated cells were analyzed by flow cytometry followed by sorting, using lectins WGA and LEL as markers. The cells' morphology was assessed using cytospinning followed by diff quick staining. Results: We observed a heterogeneous cell population in HK and PI. A high number of leucocytes was isolated from HK (lymphocytes, neutrophils, and monocytes); while in PI the number of leucocytes was scarce. WGA and LEL markers failed to distinguish the cell populations. Based on their morphology, the adherent cells seemed to be enriched in monocytes and/or macrophages. Conclusions: The development of protocols for isolation and culturing of immune cells can be used as a steppingstone for further studies on fish immunity.

Keywords: European seabass; immune cells; in vitro tools

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